

Developing Peptide MRM-based Assays for Cardiovascular Biomarker Proteins in Plasma Using a Hybrid Triple Quadrupole Linear Ion Trap Mass Spectrometer

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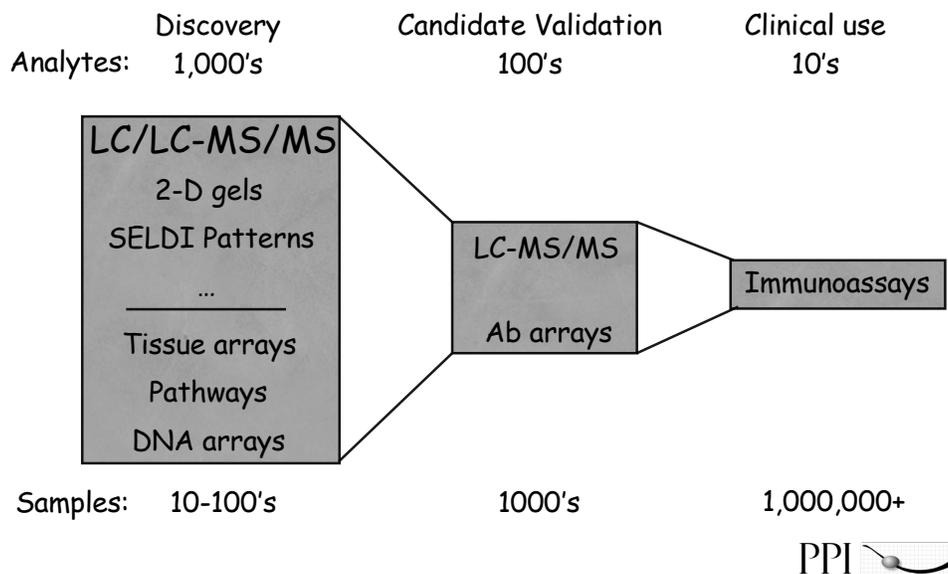
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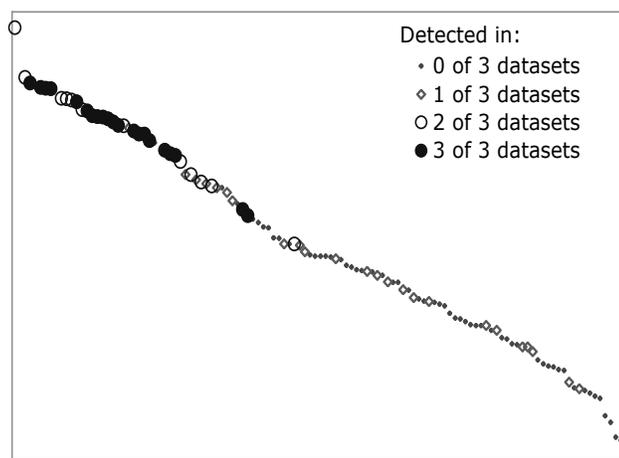
ASMS June 2005 San Antonio



Biomarker Pipeline



Plasma Proteome Discovery Platforms Have Limited Sensitivity and Lack Comprehensiveness



From: Candidate-Based Proteomics in the Search for Biomarkers of Cardiovascular Disease, Leigh Anderson, *J. Physiol.*, 563.1, 23-60 (2005)



Role for Candidate-Based Proteomics

- Multiplexed specific assays (candidate approach) can address three important drawbacks of biomarker discovery platforms for the middle pipeline stage (verification/validation):
 - Throughput
 - Sensitivity
 - Coverage (range of analytes)
- Candidate-based approaches forego most of the potential for discovery of new biomarkers, but retain some ability to "discover" optimal multi-analyte panels.
- Our purpose is to develop an MS-based candidate approach for high-throughput biomarker verification/validation in human plasma and serum



Published List of 177 CVD Candidates

Candidate-Based Proteomics in the Search for Biomarkers of Cardiovascular Disease,

Leigh Anderson, J. Physiol 563.1, 23-60 (2005)

Table 1. A table of 177 candidate markers of cardiovascular disease (CVD) and stroke, assembled through literature search

Name	Accession	Normal concentration (pg ml ⁻¹)	Source for concentration	Reason	Coagulation	Lipo-protein	Acute Phase
1 activin A	P08476	6.0E +02	(Eliar-Geva et al. 2001)	Released by heparin from vascular endothelium (Phillips et al. 2000)			
2 adiponectin (ADPN)	Q15848	4.8E +06	(Mallamaci et al. 2002)	Higher levels in essential hypertensives (Mallamaci et al. 2002)			
3 albumin	P02768	4.1E +10	(Specialty Laboratories, 2001)	Negative acute phase reactant, lower levels associated with increased risk of cardiovascular mortality (Shaper et al. 2004)			+
4 aldolase C	P09972	4.0E +03	(Asaka et al. 1990)	A more specific and sensitive marker of cerebrovascular diseases than aldolase A (Asaka et al. 1990)			
5 alpha 2 antiplasmin (alpha 2 AP)	P08697	7.0E +07	Progen test insert	An important regulator of the fibrinolytic system		+	
6 alpha 2 macroglobulin (alpha 2 M)	P01023	1.8E +09	(Specialty Laboratories, 2001)	Major plasma protease inhibitor			
7 alpha(1)-anti-chymotrypsin (ACT)	P01011	4.2E +07	(Putnam, 1975)	Major plasma protease inhibitor			+



In Silico Selection of MRM Peptides

One or more tryptic peptides used as quantitative surrogates for the protein ("monitor" peptide concept)

"Inside every bad protein there is at least one good peptide"

Began with 29,155 peptides from "mature" protein forms (21,609 unique)

Downloaded SP annotation & computed parameters

Looked for occurrence in Pounds exptl data set

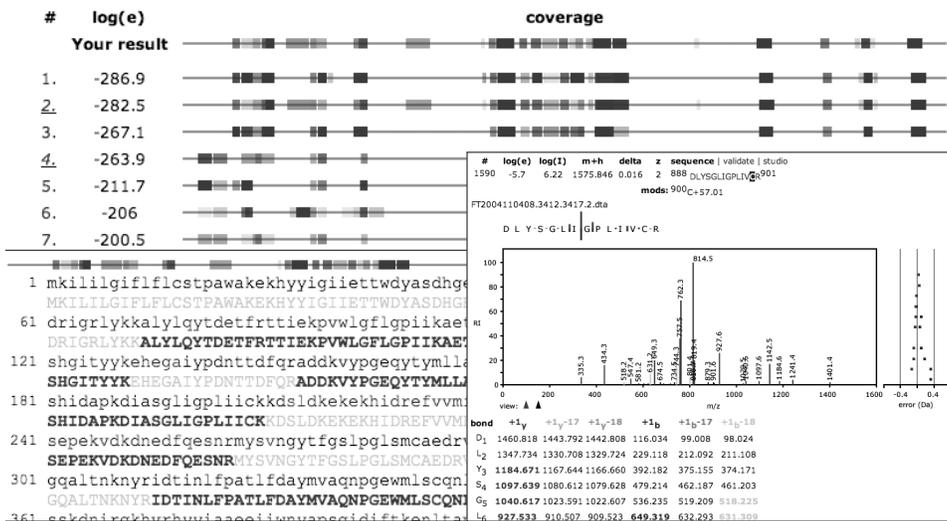
Ranked peptides on composite index of desirable properties

Term	Explanation
asn * -2 +	Susceptible to deamidation, which would change the mass of a peptide containing it.
gln * -2 +	
met * -3 +	Susceptible to oxidation, which would change the mass of a peptide containing it.
trp * -3 +	
cys * -10 +	
pro * 5 +	Produces enhanced peptide structure and can improve immunogenicity
kp * 2 +	Results in an additional positive charge in a peptide
rp * 2 +	
dp * 5 +	Introduces a site of efficient gas-phase fragmentation and improve detection
chymo_sites * -3 +	Peptide more likely to be degraded by chymotrypsin activity
occurrences_in_protein * 2 +	Multiply occurring peptide would give better detection
carbohydr * -50 +	Glycosylation of a peptide changes its mass and decreases MS detectability
mod_res * -50 +	Any documented chemical modification of a residue in the peptide changes its mass
conflicts * -20 +	Any potential sequence errors in the peptide decrease its usefulness as an MS analyte
variants * -10 +	Any genetic variants in the peptide decrease its usefulness as an MS analyte by restricting its use to a subset of patients
sign(calc_mass-800)*100 +	Applies a negative value to mass less than 800
sign(1800-calc_mass)*100 ;	Applies a negative value to mass greater than 2,000
sign(detect - 0.0001) * 10 + detect * 20 +	Gives 10 points for detection in the Pounds experimental MS/MS data if the peptide is the most frequently detected for its protein (see aforementioned index of detectability)



Beavis' GPM Useful in MRM Design

gpmDB current contains **507** entries for *ENSP0000264613*
Your result is compared against the 20 best coverage patterns.



MRM-Triggered IDA to Develop Peptide MRM Transitions

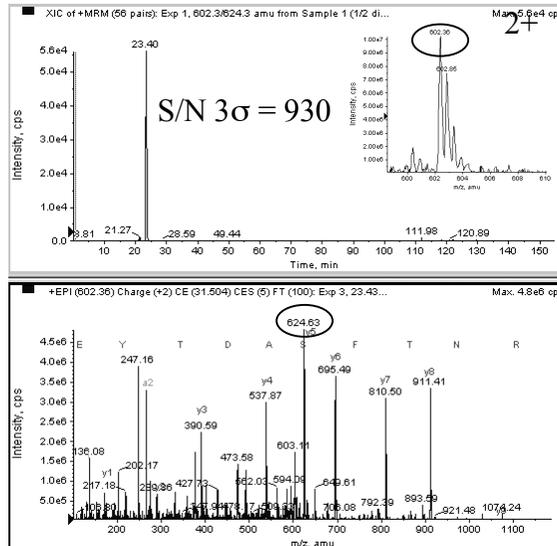
(workflow now called MIDAS: MRM-Initiated Detection and Sequencing)

Protein Sequence: MKLILGLFLCSTPAWAKEKHYIGIETTWDYASDHGKLLISV...
Enzyme: Trypsin
Missed Cleavages: 0
Fixed Modifications: 0
Variable Modifications: 1
Acquisition Method Details: Modification: (none) (QWERTYIPASDFGHKLVN)
AA Residue: QWERTYIPASDFGH
Positive Fragments: [y] > precursor [0], [2] > precursor [0]
Negative Fragments: [Not set] [0], [Not set] [0]

- Use script to build de novo MRM's to some plasma proteins
- MRM triggered IDA to confirm ID
- Use first and second y ion above precursor mass



Example Evidence for Ceruloplasmin (8 peptides found)

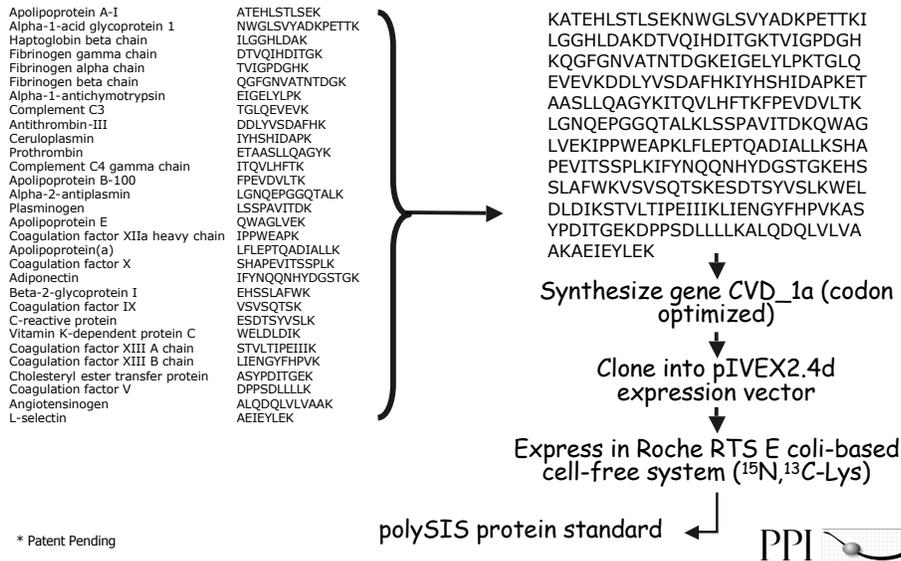


MRM's Selected for polySIS CVD_1 Peptides

Order	Protein	Peptide	Unlabeled		Labeled		CE
			Q1	Q3	Q1	Q3	
1	Apolipoprotein A-I	ATEHLSTLSEK	405.88	664.35	408.54	672.35	25
2	Alpha-1-acid glycoprotein 1	NWGLSVYADKPETTK	570.29	1052.53	575.61	1068.53	35
3	Haptoglobin beta chain	ILGGHLDAK	462.26	697.36	466.26	705.36	31
4	Fibrinogen gamma chain	DTVQIHDTGK	409.54	670.35	412.21	678.35	23
5	Fibrinogen alpha chain	TVIGPDGHHK	462.25	723.38	466.24	731.38	25
6	Fibrinogen beta chain	QGFQNVATNTDGK	654.80	706.34	658.80	714.34	30
7	Alpha-1-antichymotrypsin	EIGELYLPK	531.29	819.46	535.29	827.46	26
8	Complement C3	TGLQEVVVK	501.77	731.39	505.77	739.39	25
9	Antithrombin-III	DDLQVSDAFHK	437.21	803.40	439.87	811.40	27
10	Ceruloplasmin	IYHSHIDAPK	394.20	767.40	396.90	775.40	25
11	Prothrombin	ETAASLLQAGYK	626.33	879.49	630.33	887.49	32
12	Complement C4 gamma chain	ITQVLHFTK	362.88	645.37	365.55	653.37	30
13	Apolipoprotein B-100	FPEVDLTK	524.28	803.45	528.28	811.45	25
14	Alpha-2-antiplasmin	LGNQEPGGQTALK	656.84	771.44	660.84	779.44	40
15	Plasminogen	LSSPAVITDK	515.79	743.43	519.79	751.43	25
16	Apolipoprotein E	QWAGLVEK	465.75	616.37	469.75	624.37	28
17	Coagulation factor XIIIa heavy chain	IPPWEAPK	469.25	727.39	473.25	735.39	27
18	Apolipoprotein(a)	LFLEPTQADIALLK	786.45	1069.63	790.45	1077.63	33
19	Coagulation factor X	SHAPEVITSSPLK	455.92	632.36	458.58	640.36	28
20	Adiponectin	IFYNQNHVDGSGTK	591.27	727.33	593.94	735.33	30
21	Beta-2-glycoprotein I	EHSSLAFWK	582.77	838.45	586.77	846.45	23
22	Coagulation factor IX	VSVSQTSK	418.22	736.38	422.22	744.38	24
23	C-reactive protein	ESDTSYVSLK	564.77	696.39	568.77	704.39	29
24	Vitamin K-dependent protein C	WELDLDIK	516.27	716.42	520.27	724.42	25
25	Coagulation factor XIII A chain	STVLTIPKIIK	663.91	712.46	667.91	720.46	26
26	Coagulation factor XIII B chain	LIENGYFHPVK	439.57	847.45	442.23	855.45	25
27	Cholesteryl ester transfer protein	ASYPDITGEK	540.76	759.39	544.76	767.39	30
28	Coagulation factor V	DPPSDLLLLK	555.82	898.56	559.82	906.56	30
29	Angiotensinogen	ALQDQLVVAAK	634.88	956.58	638.88	964.58	34
30	L-selectin	AEIEYLEK	497.75	794.43	501.75	802.43	22



Efficient Production of Stable Isotope-labeled Standards (SIS) as polySIS*



Absolute Protein Quantitation in Relation to polySIS* Peptide Standards

polySIS CVD_1 (lys-labeled standard)

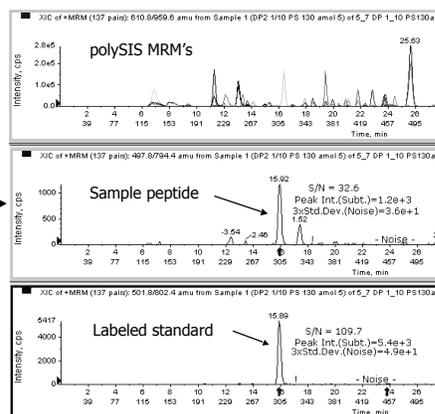
ATEHLSTLSEKNWGLSVYADKPETTKILGG HLDKADTVQIHDITGKTIVIGPDGKHQKQFG NVATNTDGEIGELYLPKTLGLQEVVKKDDL YVSDAFHKIYHSHIDAPKETAASLLQAGYKI TQVLHFTKFEVDVLTQLGNQEPGGQTALK LSSPAVITDKQWAGLVEKIPPEWAPKLFLE PTQADIALLKSHAPEVITSSPLKIFYNQNH YDGSGTKEHSSLAFWKVSVSQTSKESDTS YVSLKWELDLDIKSTVLTPEIILKIENGYF HPVKASYPDITGEKDPSPDLLLLKALQDQL VLVAAKAEIYLEK

sL-Selectin (in sample)

WTYHYSEKPMNWQRARRFRCDNYDNLVAI QNKAEIYLEKTLPFSRSYYWIGIRKIGGI WTWVGTNKSLEEAENWGDGEPNNKKNK EDCVEIYKRNKDAGKWDDACHKLKAAAL CYTASCQPWCSGSHGECVEIINNYTNCND VGYGYPQCQFVIQCEPLEAPELGTMDCTHP LGNFSFSQCAFSCSEGTNLGTGIEETTCGPF GNWSSPEPTCQVIQCEPLSAPDLGIMNCSS PLASFSFTSACTFICSEGTGELIGKKTICESS GIWNSNPICQKLDKFSMIKEGDYD

Digest

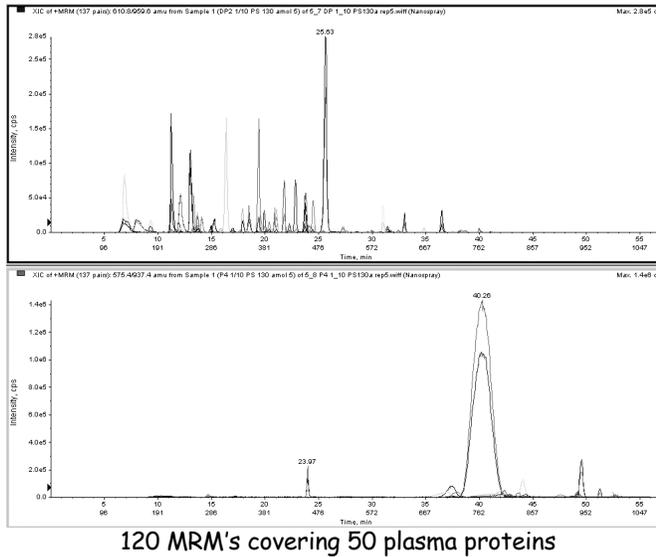
LC-TQMS of AEIYLEK



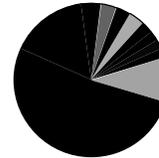
* Patent Pending



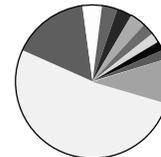
Subtraction of Top 6 Proteins (albumin, IgG, IgA, haptoglobin, transferrin and antitrypsin) Using Agilent MARS Column



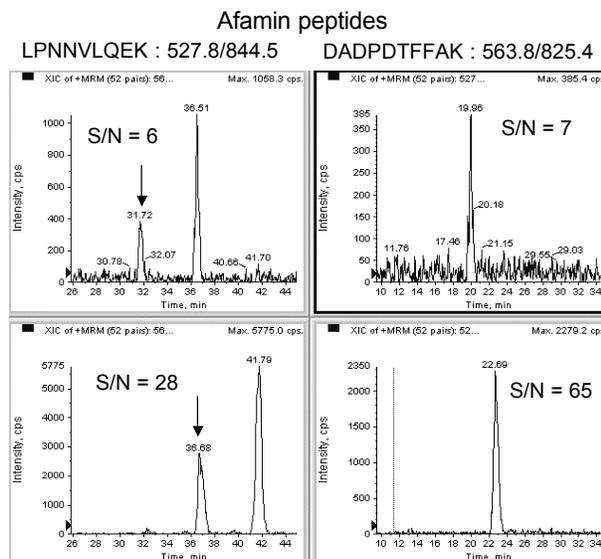
Depleted Plasma
Digest of 0.01 uL
plasma on column



Whole Plasma
Digest of 0.01 uL
plasma on column

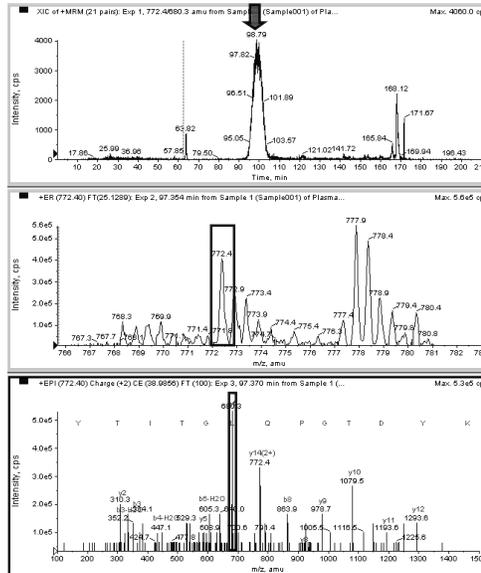


Immunosubtraction of Top 6 Proteins Yields 5-9-fold Improved S/N

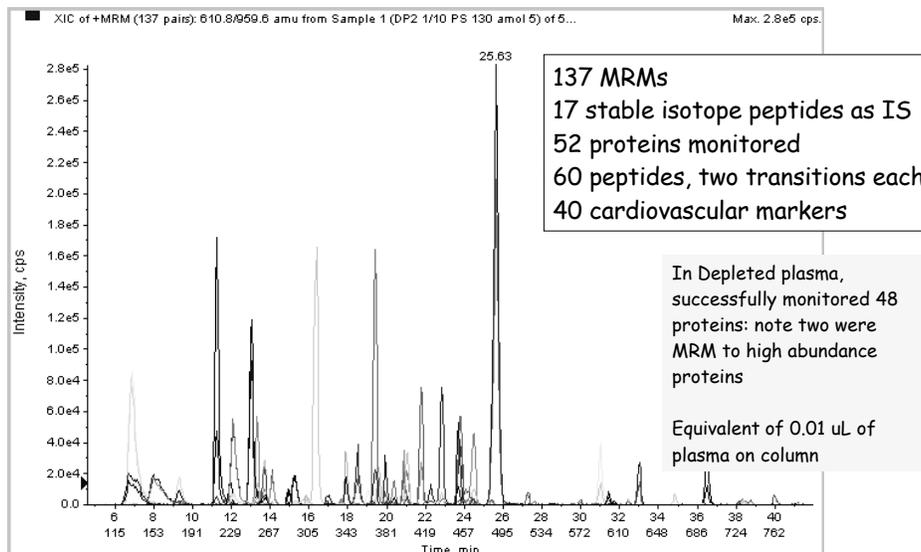


Fibronectin
SYTITGLQPGTDYK
1543.8

Fibronectin, a protein of much lower normal abundance (1.4µg/ml) could be measured using peptide SYTITGLQPGTDYK (selected *in silico*) using the transition 772.4/680.3 with S/N of 170, suggesting an LLOQ of ~100ng/ml.



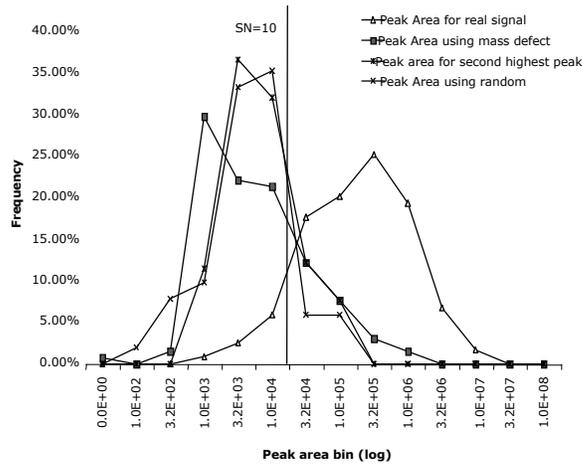
Current MRM Method



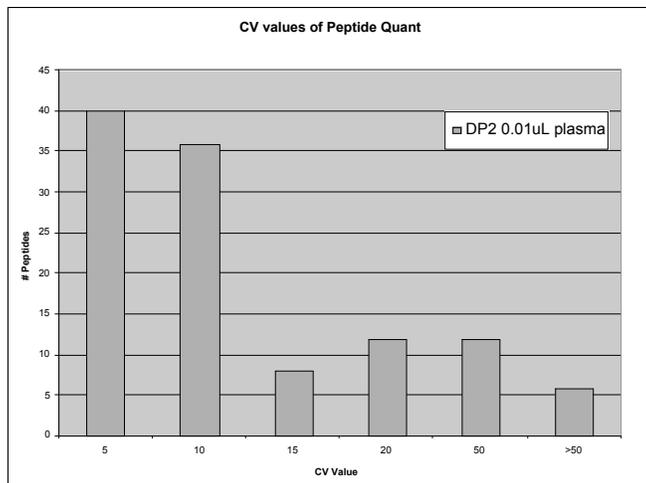
Designed vs "Random" MRM's

Occupancy of Plasma MRM Space Is Low

- Apex of the random vs real distributions are >2 orders of magnitude apart
- If we use a cutoff of peak areas for real signal at 2×10^4 , then only 10% of MRM channels will contain random signal



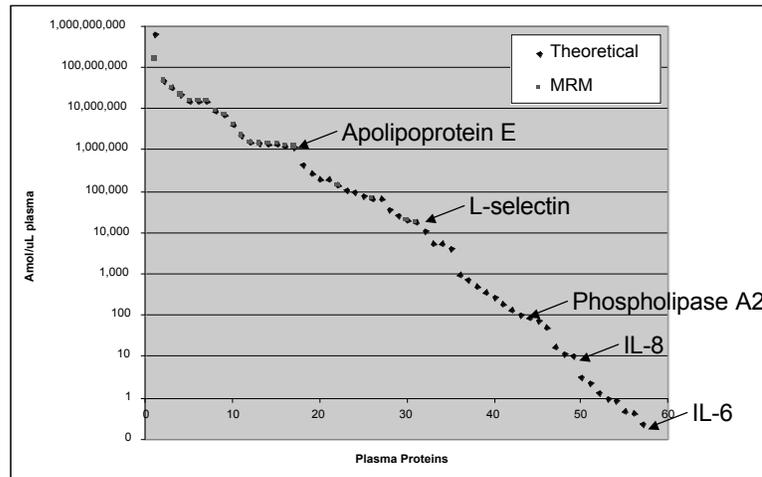
Reproducibility of MRM Panel in Depleted Plasma



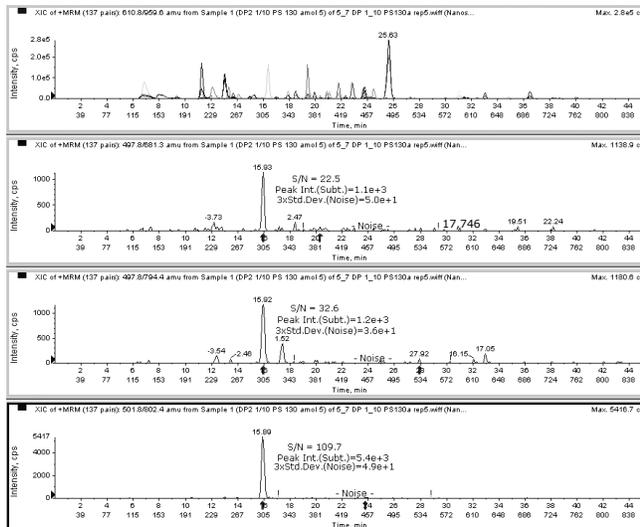
Avg CV with IS 7%
 Avg CV overall for good peaks 9.5%



Dynamic Range of Current Method in Depleted Plasma: ~4.5 Orders



Dynamic Range of Current Method Detection of L-selectin



Peak area ratio for the L-selectin monitor peptide (AEIEYLEK 497.8/794.4) and SIS (501.8/802.4) standard was 0.216.

Given 1,300 amol SIS loading (one copy of this peptide per molecule of intact L-selectin) = 280 amol L-selectin per 0.01ul injection.

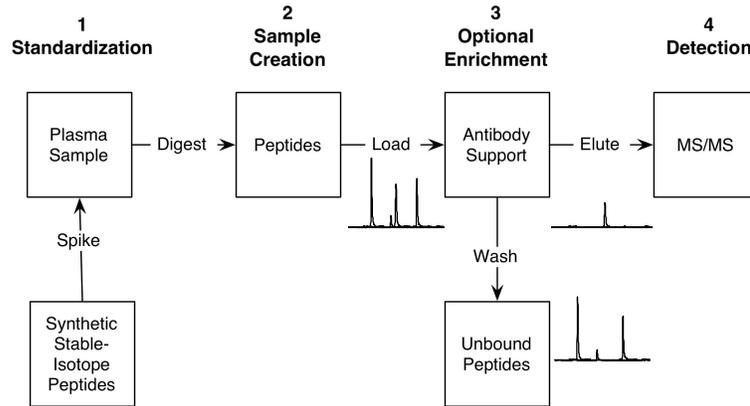
Molecular weight of plasma L-selectin is ~35,000, giving a measured concentration of 980 ng/ml vs published normal value of 670 ng/ml.

The dynamic range between albumin (~55mg/ml), and L-selectin is ~4.5 orders of magnitude



SISCAPA*: A Method Combining The Specificity of MS Detection with Sensitivity of Antibody Capture

(SISCAPA = Stable Isotope Standards with Capture by Anti-Peptide Antibody)

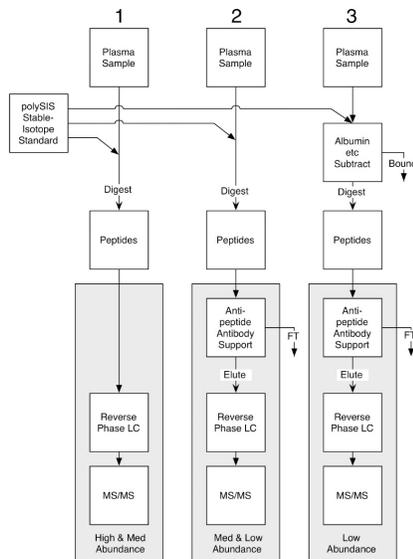


* patent pending

Mass Spectrometric Quantitation of Peptides and Proteins Using Stable Isotope Standards and Capture by Anti-Peptide Antibodies (SISCAPA). Anderson, N.L., et al, Journal of Proteome Research, 3: 235-44 (2004).



Strategies for Progressive Increases in Sensitivity of LC-MS Peptide Quantitation



Our approach generates a coherent, layered series of methods based on a single analytical platform (TQMS), with an explicit sensitivity vs cost tradeoff



Conclusions

- Large numbers of candidate biomarkers already exist to jumpstart verification/validation, and from which improved panels could be constructed
- MRM assays of monitor peptides offer a potentially rapid path to verification/validation with less cost and effort than sandwich immunoassays
- Optimal MRM design makes use of both *in silico* and experimental data
- Current MRM's appear able to access the top 5 logs of plasma protein abundance, and cover everything visible on a plasma 2-D gel
- Novel paths to creation of internal standards (e.g., polySIS proteins) can facilitate assays development
- Marker panel development and validation in large sample sets (e.g., epidemiological studies) now appears feasible



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 Drake, Gerry Hoehn, Clinical
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www.plasmaproteome.org

Recent Relevant Papers

- The human plasma proteome: History, character, and diagnostic prospects.** Anderson, N.L. and Anderson, N.G., *Molecular and Cellular Proteomics*, 1.11, 845-867 (2002)
- The human serum proteome: Display of nearly 3700 chromatographically separated protein spots on two-dimensional electrophoresis gels and identification of 325 distinct proteins.** Pieper, R., et al *Proteomics* 3(7): 1345-64. (2003).
- Multi-component immunoaffinity subtraction chromatography: An innovative step towards a comprehensive survey of the human plasma proteome.** Pieper, R., Su, Q., Gatlin, C. L., Huang, S. T., Anderson, N. L., Steiner, S. *Proteomics* 3(4): 422-32 (2003).
- Therapeutic potential of the plasma proteome.** Lathrop, J.T., Anderson, N.L., Anderson, N.G., and Hammond, D.J. *Current Opinion in Mol. Therapeutics* 5:250-257 (2003).
- Mass Spectrometric Quantitation of Peptides and Proteins Using Stable Isotope Standards and Capture by Anti-Peptide Antibodies (SISCAPA).** Anderson, N.L., Anderson, N.G., Haines, L.R., Hardie, D.B., Olafson, R.W., and Pearson, T.W. *Journal of Proteome Research*, 3: 235-44 (2004).
- NHLBI Clinical Proteomics Working Group Report.** Granger, C.B., Van Eyk, J.E., Mockrin, S.C., and Anderson, N.L., on behalf of the Working Group Members. *Circulation* 109: 1697-703 (2004).
- Candidate-Based Proteomics in the Search for Biomarkers of Cardiovascular Disease,** Leigh Anderson, J. *Physiol.*, 563.1, 23-60 (2005)

